# **Tetrodotoxins Secretion and Voltage-Gated Sodium Channel Adaptation in Ribbon Worm** Kulikovia alborostrata (Takakura, 1898) (Nemertea)

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Nemertea is a phylum of marine worms, counting more than 1300 species, most of which are active predators. Many species of these worms bear various toxins, including tetrodotoxin (TTX) – a potent low molecular weight neurotoxin of bacterial origin.

Despite the more than 30 years of studying TTX in nemerteans, many



questions regarding its functions, mechanisms ensuring its accumulation and usage remain unclear. For many TTX-secreting animals, the function of the toxin as the predators' deterrent was suggested. The realization of this function in ribbon worms supposes the recovery of TTX in secreting cells through migration from the tissues of the internal environment. In the current research, using 17 specimens of the ribbon worm Kulikovia alborostrata, we studied the dynamics of TTXs concentration in the secretion produced at different time intervals and toxins localization at different stages of the excretion process. To accumulate TTXs and specifically use them as antipredator defense or for prey immobilizing during hunting, animals should have molecular mechanisms ensuring resistance to the toxin. The resistance mechanisms known for some TTX-bearing animals represent mutations in TTX targets – voltage-gated sodium (NaV) channels in the region of the selective filter. In the current research, for the first time, a search for the amino acid substitutions, leading to a decrease of the affinity of the NaV1 channel to TTX in nemerteans, was performed.

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TTXs removal	TTXs recovery in secreting structures	Nav adaptations			
Repeated stimulations at short intervals for the depletion of the TTXs from the secretion structures	Investigation of recovery of the TTXs in the nemertean secreting structures during long-term captivity without food after TTXs removal	Search for substitutions in the P-loop regions of Nav1 channel domains I-IV that contribute to the TTX-resistance of nemerteans			
	Sanger sequencing				
Electrical pulses (2	2 s, 6V) $\bigvee$ Secretions sample $\bigvee$ Body sample	<b>V</b> Sample processing			

Figure 1. Scheme of the experimental design for studies of tetrodotoxins (TTXs) secretion and Nav1 adaptation in the ribbon worm Kulikovia alborostrata

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N	e. of specimen	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
Day 1 —	TTX	+	+	+	-	+	-	+	-	-	-	-	_	-	-	-	_	-
	5,6,1 ltrideoxyTTX	0.675	0.264	0.229	- (	-	-	-	-	-	-	-	-	0.144	. –	-	_	-
Day 2 —	TTX	0.340	0.357	+	0.556	i +	+	+	+	+	-	+	+	-	-	-	_	+
	5,6,1 ltrideoxyTTX	1.905	3.171	0.156	6.244	0.399	-	1.690	0.774	-	0.440	_	0.835	_	-	-	_	0.19
Secretion (TTXs, Day 3 —	TTX	-	-	-	-	-	-	-	+	-	-	-	-	-	-	+	_	-
	5,6,1 ltrideoxyTTX	-	-	-	-	-	-	-	-	-	-	_	0.287	0.115	-	0.093	_	-
ng/g) Day 30	0 TTX	+	+	-	0.306	-	-	-	-	-	+	-	+	+	-	+	_	-
	5,6,1 ltrideoxyTTX	-	+	+	-	0.338	+	0.862	0.413	+	0.253	0.178	80.547	0.140	+	0.068	_	+
Day 21	$_{1}$ TTX	+	+	+	+	+	+	+	+	0.305	· +	+	+	+	+	+	+	+
	5,6,1 $trideoxyTTX$	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Body (TTXs, ng/	TTX	+	+	+	1.166	0.492	0.146	5 +	+	1.286	+	0.306	50.441	+	+	+	+	+
	5,6,1 ltrideoxyTTX	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	_	-

#### **TTXs removal and recovery in secreting structures**

Table 1. Tetrodotoxin (TTX) and 5,6,11-trideoxyTTX in extracts of nemertean Kulikovia alborostrata. +: <limit of quantification (0.6 ng/mL of extract); -: not detected.

### **TTXs secretion**

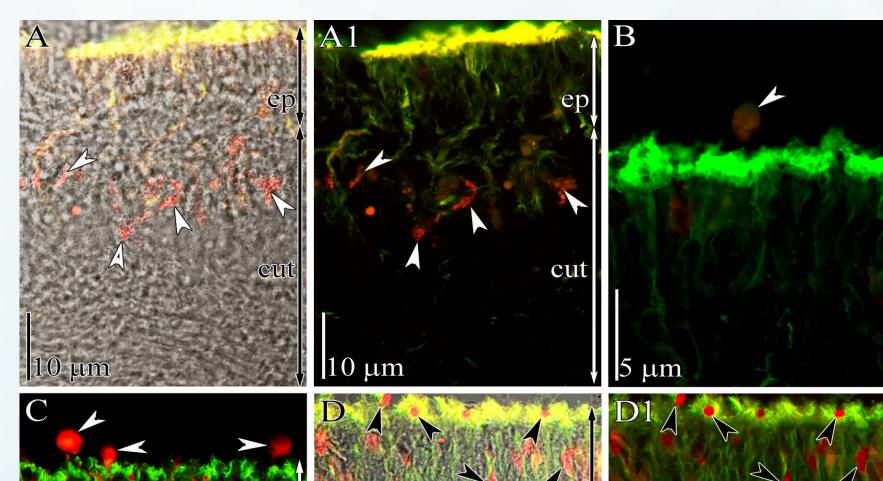


Figure 2. Confocal laser scanning micrographs (Z-projections) of transverse sections of the body

wall of the intact (A,B) and stimulated (C,D) specimens of Kulikovia alborostrata. Red: tetrodotoxin-like immunoreactivity; green: cilia and cytoskeleton, acetylated tubulin immunoreactivity.

(A,A1) Panoramic view showing subepidermal gland cells with toxin-positive granules (arrowheads). (B) TTX-positive spherical secretion (arrowhead) above the apical surface of the epidermis. Panoramic vep view showing TTX-positive spherical secretions (arrowheads) above the apical surface of the epidermis. (D,D1) Panoramic view of integument showing subepidermal gland cells with TTX-positive granules (white arrowheads) and their epidermal extensions with TTX-positive granules (black arrowheads). Legends: cut, cutis; ep, epithelium.

#### **TTXs** adaptation

	Domain I	Domain II	Domain III	Domain IV
	404	758	1239	1534
Homo sapiens NaV1.4	TQ <mark>DY</mark> W <mark>E</mark> NLFQ	CG <mark>E</mark> WI <mark>E</mark> T	VAT <mark>FK</mark> GWMDI	TS <b>AG</b> W <mark>D</mark> GLLN
<i>Takifugu rubripes</i> NaV1.4b	TQ <b>D</b> FW <mark>E</mark> NLFQ	CG <mark>e</mark> wi <mark>e</mark> S	VAT <mark>F</mark> KGW <b>T</b> DI	TS <b>A<mark>G</mark>WD</b> GLLS
<i>Takifugu rubripes</i> NaV1.6b	??D <mark>Y</mark> WE <mark>G</mark> ???	?? <mark>e</mark> wiet	??? <mark>?K</mark> GWMD?	?? <b>A<mark>G</mark>WDG???</b>
Thamnophis sirtalis NaV1.4	TQ <mark>D</mark> YW <mark>E</mark> NLFQ	CG <mark>e</mark> wiet	VAT <mark>F</mark> KGWMDI	TS <b>AGW<mark>NV</mark>LLN</b>
<i>Taricha granulosa</i> NaV1.3	TQ <b>DA</b> W <mark>E</mark> NLYQ	CG <mark>e</mark> wie <mark>s</mark>	VAT <mark>F</mark> KGWMDI	TS <b>A<mark>G</mark>WDGLLA</b>
Notospermus geniculatus NaV1	TQ <mark>D</mark> YW <mark>E</mark> NLYQ	CG <mark>e</mark> wi <mark>e</mark> S	VAT <mark>F</mark> KGW <b>T</b> EI	TS <b>A<mark>G</mark>W<mark>N</mark>YVLN</b>
Kulikovia alborostrata NaV1	TQ <mark>D</mark> YWE <mark>G</mark> VYH	CG <mark>e</mark> wie <mark>s</mark>	VAT <mark>FK</mark> GW <b>T</b> EI	TS <b>AG</b> W <mark>N</mark> SVLD

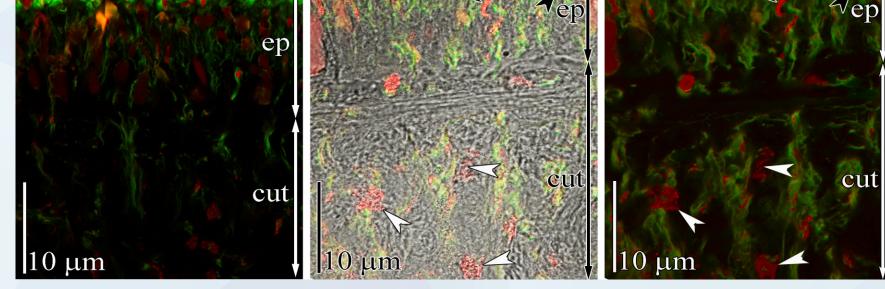


Figure 3. Sequence alignment of P-loop regions of Nav channels domains I-IV of Homo sapiens(TTX-sensitive channel), TTX-resistant animals (Takifugu rubripes, Thamnophis sirtalis, Taricha granulosa), and Kulikovia alborostrata, studied herein. The four key amino acids constituting the selectivity region—aspartate-glutamate-lysinealanine (DEKA) motif are marked in blue. Amino acid substitutions associated with TTX channel resistance are marked in red. Amino acids involved in TTX binding and coordination are marked in yellow. Accession numbers or references: H. sapiens Nav1.4: P35499; T. rubripes Nav1.4b: Q2XVR6; T. rubripes Nav1.6b; T. granulosa Nav1.3: A0A6G9W273; T. sirtalis Nav1.4: A0A1W5T2B2.

#### The studies have shown:

- A low rate of TTX recovery in the nemertean secreting structures; after 30 days, TTX was recovered in only one-third of the studied ribbon worms. This leads us to suggest a low probability of targeted usage of TTX by the toxic nemertean K. alborostrata as a repellent through its secretion in mucus.
- The TTX analogue 5,6,11-trideoxyTTX, on the contrary, fully migrated from the body wall into the secreting cells and was completely lost through the release of secretion after 30 days of the experiment, which may indicate the specificity of its transfer in response to stimulation and possible targeted usage.
- The sequences of the P-loop regions of Nav1 channel domains I–IV of all 17 studied specimens are identical and have amino acid substitutions, which were shown for TTX-resistant organisms, and, according to the literature data, can contribute to TTX resistance.